

09/994,657

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DB=USPT,JPAB,EPAB,DWPI; PLUR=YES; OP=ADJ

L4

L3 and (time\$1 or period\$1)

23

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23

L3L2

L1 and volt

53

L2

DB=DWPI,USPT,EPAB,JPAB; PLUR=YES; OP=ADJ

L1

electr\$7 near5 bacterial

435

L1

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- ☐ 1. 6520950. 08 May 00; 18 Feb 03. Method of electroporation-enhanced delivery of active agents. Hofmann; Gunter A., et al. 604/503; A61M025/00.
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- ☐ 2. 6399861. 23 May 95; 04 Jun 02. Methods and compositions for the production of stably transformed, fertile monocot plants and cells thereof. Anderson; Paul C., et al. 800/320.1; 800/275 800/288 800/293 800/301 800/302 800/303. A01H005/00 C12N005/04.
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- ☐ 3. 6350934. 12 Jul 96; 26 Feb 02. Nucleic acid encoding delta-9 desaturase. Zwick; Michael G., et al. 800/281; 435/320.1 435/412 435/419 435/469 435/470 536/23.2 536/23.6 800/278 800/286 800/287 800/292 800/293 800/294 800/300 800/320.1. C12N005/04 C12N015/29 C12N015/82 A01H005/00.
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- ☐ 4. 6335161. 25 Feb 98; 01 Jan 02. Release of intracellular material and the production therefrom of single stranded nucleic acid. Martin; Sophie E.V., et al. 435/6; 435/91.2 436/94. C12Q001/68 C12P019/34 G01N033/48.
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- ☐ 5. 6329574. 24 Jul 98; 11 Dec 01. High lysine fertile transgenic corn plants. Lundquist; Ronald C., et al. 800/300.1; 800/278 800/287 800/288 800/293 800/320.1. C12N015/00 A01H001/06 A01H004/00.
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- ☒ 6. 6302874. 13 Jul 99; 16 Oct 01. Method and apparatus for electrically assisted topical delivery of agents for cosmetic applications. Zhang; Lei, et al. 604/522; 604/501. A61M031/00.
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- ☒ 7. 6103235. 15 Apr 97; 15 Aug 00. Methods of inducing immune tolerance using immunotoxins. Neville; David M., et al. 424/183.1; 424/184.1. A61K039/395 A61K039/00.
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- ☐ 8. 6025545. 15 May 95; 15 Feb 00. Methods and compositions for the production of stably transformed, fertile monocot plants and cells thereof. Lundquist; Ronald C., et al. 800/300.1; 536/23.1 536/24.1 800/298 800/300 800/320.1. A01H001/06 A01H004/00 C12M015/00.
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- ☐ 9. 5990390. 27 Mar 96; 23 Nov 99. Methods and compositions for the production of stably transformed, fertile monocot plants and cells thereof. Lundquist; Ronald C., et al. 800/302; 536/23.71 800/265 800/268 800/320.1. A01H005/00 A01H004/00 A01H001/20 C12H005/04.
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- ☒ 10. 5989846. 06 Jun 95; 23 Nov 99. Assays to identify inducers of plant defense resistance. Klessig; Daniel Frederick, et al. 435/27; 435/184 435/28. C12Q001/30 C12Q001/28.
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L2 ANSWER 1 OF 4 MEDLINE
AN 90073642 MEDLINE
DN 90073642 PubMed ID: 2686636
TI A rapid and efficient procedure for transformation of intact Saccharomyces cerevisiae by electroporation.
AU Simon J R; McEntee K
CS Department of Biological Chemistry, UCLA School of Medicine 90024.
NC GM 38456 (NIGMS)
SO BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, (1989 Nov 15) 164 (3) 1157-64.
Journal code: 0372516. ISSN: 0006-291X.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 198912
ED Entered STN: 19900328
Last Updated on STN: 19970203
Entered Medline: 19891228
AB A rapid and efficient procedure is described for transforming Saccharomyces cerevisiae using **electroporation** to render intact cells **permeable** to DNA. The technique uses relatively low voltages and is particularly sensitive to low concentrations of plasmid DNA. At the highest voltage used (400 volts), the frequency of transformation increased with the amount of plasmid DNA

between 25 ng and 100 ng. At higher concentrations of DNA (1-1.5 micrograms) electroporation yielded one-third to one-half the number of transformants obtained with a standard lithium acetate pretreatment. Because this method requires neither pretreatment of cells nor addition of polyethylene glycol (PEG), it has several advantages over currently used transformation procedures.

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L2 ANSWER 2 OF 4 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.

AN 1990:48268 BIOSIS

DN BA89:25632

TI A RAPID AND EFFICIENT PROCEDURE FOR TRANSFORMATION OF INTACT *SACCHAROMYCES-CEREVISIAE* BY ELECTROPORATION.

AU SIMON J R; MCENTEE K

CS DEP. BIOLOGICAL CHEM., UCLA SCH. MED., LAB. BIOMED. ENVIRONMENTAL SCIENCES, 900 VETERAN AVE., LOS ANGELES, CALIF. 90024.

SO BIOCHEM BIOPHYS RES COMMUN, (1989) 164 (3), 1157-1164.

CODEN: BBRCA9. ISSN: 0006-291X.

FS BA; OLD

LA English

AB A rapid and efficient procedure is described for transforming *Saccharomyces cerevisiae* using **electroporation** to render intact **cells permeable** to DNA. The technique uses relatively low voltages and is particularly sensitive to low concentrations of plasmid DNA. At the highest voltage used (400 volts), the frequency of transformation increased with the amount of plasmid DNA between 25 ng and 100 ng. At higher concentrations of DNA (1-1.5 .mu.g) electroporation yielded one-third to one-half the number of transformants obtained with a standard lithium acetate pretreatment. Because this method requires neither pretreatment of cells nor addition of polyethylene glycol (PEG), it has several advantages over currently used transformation procedures.

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L2 ANSWER 3 OF 4 CAPLUS COPYRIGHT 2003 ACS

AN 1976:126439 CAPLUS

DN 84:126439

TI Electrolytic cell for inactivation and destruction of pathogenic material

IN Shaffer, Peter T. B.

PA Carborundum Co., USA

SO U.S., 6 pp.

CODEN: USXXAM

DT Patent

LA English

FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 3923629	A	19751202	US 1974-454637	19740325
	CA 1041038	A1	19781024	CA 1975-221887	19750311
	JP 50133173	A2	19751022	JP 1975-34592	19750324
PRAI	US 1974-454637		19740325		
AB	An electrolytic cell for destroying fluid-born				

pathogenic materials comprises layers of **permeable elec.** conductive material sepd. by layers of permeable elec. insulation. The conductive layers act as the cell electrodes which are connected to an a.c. source having a current potential of .apprx.0.1-20 **volts** with frequencies of .apprx.0.1-.apprx.10 Hz. A suitable filter housing surrounds the cell and is arranged so that the pathogen-contg. fluid passes through the **permeable electrode** layers of the **cell**. The pathogenic materials are subjected to the elec. potential set up between the layers and are inactivated or destroyed.

AB An **electrolytic cell** for destroying fluid-born pathogenic materials comprises layers of **permeable elec.** conductive material sepd. by layers of permeable elec. insulation. The conductive layers act as the cell electrodes which are connected to an a.c. source having a current potential of .apprx.0.1-20 **volts** with frequencies of .apprx.0.1-.apprx.10 Hz. A suitable filter housing surrounds the cell and is arranged so that the pathogen-contg. fluid passes through the **permeable electrode** layers of the **cell**. The pathogenic materials are subjected to the elec. potential set up between the layers and are inactivated or destroyed.

L2 ANSWER 4 OF 4 CAPLUS COPYRIGHT 2003 ACS

AN 1967:15861 CAPLUS

DN 66:15861

TI Preparation of thoria sols by electrodialysis

IN O'Connor, Thomas L.; Juda, Walter; McNally, Paul H.; Rosenberg, Norman W.

PA Diamond Alkali Co.

SO U.S., 10 pp.

CODEN: USXXAM

DT Patent

LA English

FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 3280011		19661018	US	19580619
AB	Fluid, aq. hydrated actinide oxide sols with a controlled and uniform particle size, neutral pH, have several advantages over solid reactor fuels. They are prepd. by putting an aq. soln. of a Th metal salt in a 1st chamber of an electrodialysis cell bounded on at least one side by a cation- permeable membrane. In a 2nd chamber on the other side of the membrane H2O is passed. An elec. current is conducted across the membrane through the solns. at room temp. The Th ions in the Th salt soln. pass through the membrane from the 1st to the 2nd chamber forming a Th oxide sol in the 2nd chamber. Thus, a soln. contg. UO2SO4 1, Th(SO4)2 100, H2SO4 5 g., in H2O to make 1 l. in the cathode compartment is subjected to d.c. (100 amp./ft.2 of an electrodialysis cell). The cell consists of 2 chambers sepd. by an anion selective membrane. After 5 hrs. of recirculating electrolysis at 85.degree. and a .apprx.5 volts d.c., the pH increases to 6.2; the actinide sol may then be withdrawn from the compartment. Av. particle size is 55 m.mu. which size is not abrasive to bends and orifices of equipment when pumped at fast rates and not small enough to form gels.				
AB	Fluid, aq. hydrated actinide oxide sols with a controlled and uniform particle size, neutral pH, have several advantages over solid reactor fuels. They are prepd. by putting an aq. soln. of a Th metal salt in a 1st chamber of an electrodialysis cell bounded on at least one side by a cation- permeable membrane. In a 2nd chamber on the other side of the membrane H2O is passed. An elec. current is conducted across the membrane through the solns. at room temp. The Th ions in the Th salt soln. pass through the membrane from the 1st to the 2nd chamber forming a Th oxide sol in the 2nd chamber. Thus, a soln. contg. UO2SO4 1, Th(SO4)2 100, H2SO4 5 g., in H2O to make 1 l. in the cathode compartment is subjected to d.c. (100 amp./ft.2 of an electrodialysis cell). The cell consists of 2 chambers sepd. by an anion selective membrane. After 5 hrs. of recirculating electrolysis at				

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=>



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